

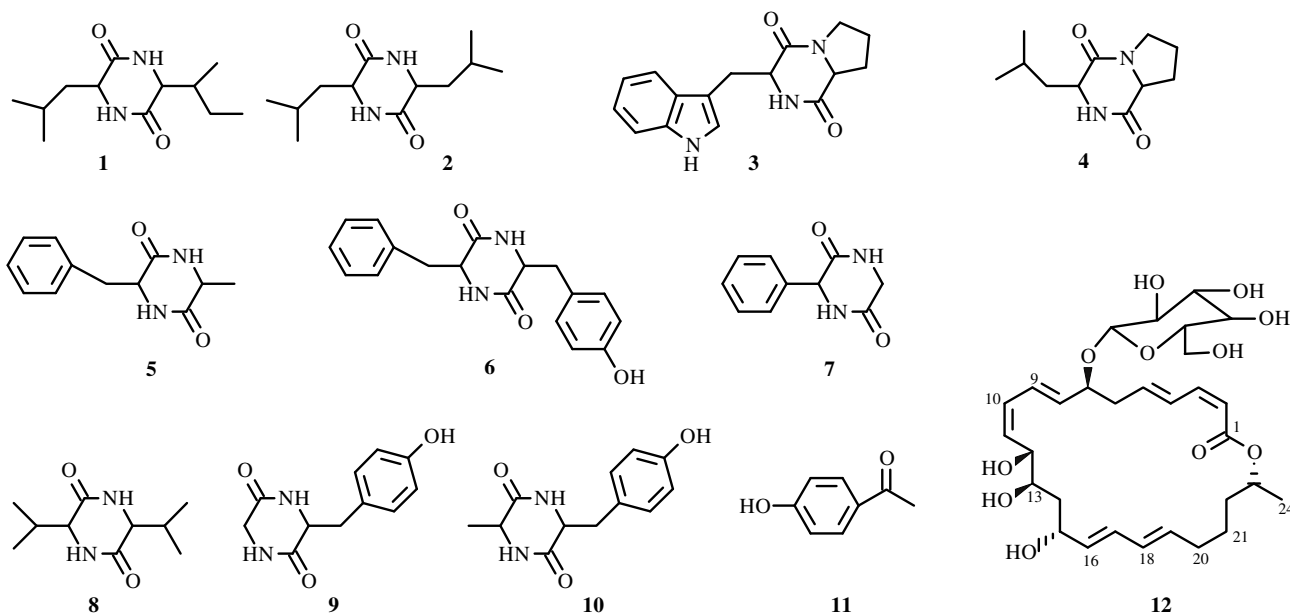
## DIKETOPIPERAZINE CONSTITUENTS OF MARINE *Bacillus subtilis*

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Marine organisms, especially marine microorganisms, have been proven to be a rich source of diverse arrays of bioactive metabolites with great potential for pharmaceutical and medical applications. Marine *Bacillus subtilis* is a common strain species, which usually produces cyclopeptide, macrolide, indole derivatives, etc. In the course of further studies, 10 diketopiperazines: cyclo(Leu-Ile) (**1**) [1], cyclo(Leu-Leu) (**2**) [2], cyclo(Trp-Pro) (**3**) [3], cyclo(Leu-Pro) (**4**) [4, 5], cyclo(Phe-Ala) (**5**) [6], cyclo(Phe-Tyr) (**6**) [2], cyclo(benzyl-Gly) (**7**) [7], cyclo(Val-Val) (**8**) [8], cyclo(Gla-Tyr) (**9**) [9], cyclo(Ala-Tyr) (**10**) [10], 1-(4-hydroxyphenyl)ethanone (**11**) [11], and macrolactin B (**12**) [12] have been isolated from the marine *Bacillus subtilis*. Among them, compound **7** is a new natural product. All the compounds have not been reported before from this species.

The strain 201721 has been derived from sea sediment of East Sea of China. It was isolated on F1 medium [Glucose (6 g), peptone (5 g), and yeast extract (1 g) were dissolved in artificial seawater (1L)] with incubation for 4 days at 28°C. Due to its chemical and morphological features as well as the 16S rDNA (GenBank accession number EF417872), the strain can be identified as the genus *Bacillus* sp.



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The entire culture broth was centrifugated at 12000 rpm for 15 min, yielding the supernatant. The supernatant was extracted with ethyl acetate 3 times. The combined extracts were evaporated in vacuo at 30°C to yield 5 g of the crude extract, which was subjected to Sephadex LH-20 column and eluted with CHCl<sub>3</sub>–CH<sub>3</sub>OH (4:1) to get five fractions, one of which was rechromatographed on middle-pressure LC with the gradient CH<sub>3</sub>OH–H<sub>2</sub>O. The fractions obtained were chromatographed on a C<sub>8</sub> semi-preparative LC (Zorbax Columns, R<sub>X</sub>-C<sub>8</sub>, 9.4 mm × 25 cm, 2 mL/min, 218 nm) using isocratic elution of CH<sub>3</sub>CN–H<sub>2</sub>O, yielding compounds **1** (2 mg), **2** (2 mg), **3** (3 mg), **4** (2 mg), **5** (1 mg), **6** (3 mg), **7** (2 mg), **8** (1 mg), **9** (5 mg), **10** (3 mg), **11** (5 mg), and **12** (3 mg).

**Cyclo(Leu-Ile) (1):** white powder, C<sub>12</sub>H<sub>22</sub>N<sub>2</sub>O<sub>2</sub>. ESI-MS: *m/z* 249.5 [M+Na]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 4.0 (1H, m, Leu<sub>α</sub>), 3.8 (1H, m, Ile<sub>α</sub>), 1.93 (1H, m, Ile<sub>β</sub>), 1.74 (1H, m, Leu<sub>β</sub>), 1.72 (1H, m, Leu<sub>β</sub>), 1.71 (1H, m, Ile<sub>γ</sub>), 1.54 (1H, m, Ile<sub>γ</sub>), 1.41 (1H, m, Leu<sub>γ</sub>), 1.2 (3H, d, Ile<sub>β</sub>-CH<sub>3</sub>), 1.19 (3H, m, Leu<sub>γ</sub>-CH<sub>3</sub>), 0.88 (3H, d, Leu<sub>γ</sub>-CH<sub>3</sub>), 0.80 (3H, d, Ile<sub>δ</sub>); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 169.58 (s, Leu<sub>CO</sub>), 168.08 (s, Ile<sub>CO</sub>), 60.07 (d, Ile<sub>α</sub>), 53.26 (d, Leu<sub>α</sub>), 44.49 (t, Leu<sub>β</sub>), 38.98 (d, Ile<sub>β</sub>), 24.44 (t, Ile<sub>γ</sub>), 24.33 (d, Leu<sub>γ</sub>), 23.29 (q, Ile<sub>β</sub>-CH<sub>3</sub>), 21.26 (q, Ile<sub>δ</sub>), 15.29 (q, Leu<sub>γ</sub>-CH<sub>3</sub>), 11.72 (q, Leu<sub>γ</sub>-CH<sub>3</sub>).

**Cyclo(Leu-Leu) (2):** white powder, C<sub>12</sub>H<sub>22</sub>N<sub>2</sub>O<sub>2</sub>. ESI-MS: *m/z* 249.3 [M+Na]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 3.9 (1H, m, Leu<sub>α</sub>), 1.8 (2H, m, Leu<sub>β</sub>), 1.7 (1H, m, Leu<sub>γ</sub>), 1.5 (3H, d, Leu<sub>γ</sub>-CH<sub>3</sub>), 1.2 (3H, d, Leu<sub>γ</sub>-CH<sub>3</sub>); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 169.12 (s, Leu<sub>CO</sub>), 53.23 (d, Leu<sub>α</sub>), 43.82 (t, Leu<sub>β</sub>), 24.09 (d, Leu<sub>γ</sub>), 23.06 (q, Leu<sub>γ</sub>-CH<sub>3</sub>), 21.00 (q, Leu<sub>γ</sub>-CH<sub>3</sub>).

**Cyclo(Trp-Pro) (3):** white powder, C<sub>16</sub>H<sub>17</sub>N<sub>3</sub>O<sub>2</sub>. ESI-MS: *m/z* 306.4 [M+Na]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ, J/Hz): 7.53 (1H, m, J = 7.8, Trp<sub>2'</sub>), 7.50 (1H, m, J = 7.2, Trp<sub>4'</sub>), 7.34 (1H, dd, J = 7.2, Trp<sub>7'</sub>), 7.12 (2H, m, J = 7.2, Trp<sub>6'</sub>), 7.03 (1H, m, J = 7.2, Trp<sub>5'</sub>), 4.36 (1H, m, Trp<sub>α</sub>), 4.00 (1H, m, Pro<sub>α</sub>), 3.52 (2H, m, Pro<sub>δ</sub>), 3.4 (1H, m, Trp<sub>β</sub>), 3.1 (1H, m, Trp<sub>β</sub>), 2.1 (1H, m, Pro<sub>β</sub>), 1.7 (2H, m, Pro<sub>β</sub>, Pro<sub>γ</sub>), 1.4 (1H, m, Pro<sub>γ</sub>); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 170.22 (s, Pro<sub>CO</sub>), 166.68 (s, Trp<sub>CO</sub>), 137.36 (s, Trp<sub>9'</sub>), 127.48 (s, Trp<sub>4'</sub>), 124.72 (d, Trp<sub>9'</sub>), 122.43 (d, Trp<sub>6'</sub>), 119.69 (d, Trp<sub>7'</sub>), 118.83 (d, Trp<sub>5'</sub>), 112.08 (d, Trp<sub>8'</sub>), 108.92 (s, Trp<sub>3'</sub>), 59.66 (d, Pro<sub>α</sub>), 55.84 (d, Trp<sub>α</sub>), 45.71 (t, Pro<sub>δ</sub>), 28.57 (t, Pro<sub>β</sub>), 27.90 (t, Trp<sub>β</sub>), 22.55 (t, Pro<sub>δ</sub>).

**Cyclo(Leu-Pro) (4):** white powder, C<sub>11</sub>H<sub>18</sub>N<sub>2</sub>O<sub>2</sub>. ESI-MS: *m/z* 233.1 [M+Na]<sup>+</sup> 211.1 [M+H]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 4.1 (1H, m, Leu<sub>α</sub>), 4.0 (1H, m, Pro<sub>α</sub>), 3.5 (2H, m, Pro<sub>δ</sub>), 2.6 (2H, m, Pro<sub>β</sub>), 1.9 (2H, m, Pro<sub>γ</sub>), 1.8 (2H, m, Leu<sub>β</sub>), 1.71 (1H, m, Leu<sub>γ</sub>), 1.1 (3H, d, Leu<sub>γ</sub>-CH<sub>3</sub>), 0.9 (3H, d, Leu<sub>γ</sub>-CH<sub>3</sub>); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 170.48 (s, Leu<sub>CO</sub>), 166.26 (s, Pro<sub>CO</sub>), 58.83 (d, Pro<sub>α</sub>), 53.4 (d, Leu<sub>α</sub>), 45.41 (t, Pro<sub>δ</sub>), 38.49 (t, Leu<sub>β</sub>), 24.54 (t, Pro<sub>β</sub>), 23.16 (d, Leu<sub>γ</sub>), 22.67 (q, Leu<sub>γ</sub>-CH<sub>3</sub>), 21.55 (t, Pro<sub>γ</sub>), 21.24 (q, Leu<sub>γ</sub>-CH<sub>3</sub>).

**Cyclo(Phe-Ala) (5):** white needle, C<sub>12</sub>H<sub>14</sub>N<sub>2</sub>O<sub>2</sub>. ESI-MS: *m/z* 241.2 [M+Na]<sup>+</sup> *m/z* 217.6 [M-H]<sup>-</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, J/Hz, δ): 7.20–7.35 (5H, m, Phe<sub>Ar</sub>), 4.27 (1H, m, J = 4.8, Phe<sub>α</sub>), 3.8 (1H, m, J = 7.2, Ala<sub>α</sub>), 3.19 (1H, m, Phe<sub>β</sub>), 3.09 (1H, m, Phe<sub>β</sub>), 0.8 (3H, d, Ala<sub>β</sub>); <sup>13</sup>C NMR (CD<sub>3</sub>OD, CD<sub>3</sub>Cl, 150 MHz): δ 167.00 (s, Phe<sub>CO</sub>), 166.26 (s, Ala<sub>CO</sub>), 134.85 (s, Phe<sub>1'</sub>), 130.00 (d, Phe<sub>3'</sub>), 128.60 (d, Phe<sub>2'</sub>), 127.30 (d, Phe<sub>4'</sub>), 56.06 (d, Phe<sub>α</sub>), 50.47 (d, Ala<sub>α</sub>), 39.62 (t, Phe<sub>β</sub>), 19.69 (q, Ala<sub>β</sub>).

**Cyclo(Phe-Tyr) (6):** white needle, C<sub>18</sub>H<sub>18</sub>N<sub>2</sub>O<sub>3</sub>. ESI-MS: *m/z* 333.4 [M+Na]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, DMSO-d<sub>6</sub>, δ, J/Hz): 9.234 (1H, s, Tyr<sub>OH</sub>), 7.836 (2H, s, Tyr<sub>NH</sub>, Phe<sub>NH</sub>), 7.287 (2H, d, J = 7.2, Phe<sub>2'</sub>), 7.203 (1H, d, J = 7.2, Phe<sub>4'</sub>), 7.043 (2H, d, J = 7.8, Phe<sub>3'</sub>), 6.841 (2H, d, J = 8.4, Tyr<sub>2'</sub>), 6.679 (2H, d, J = 8.4, Tyr<sub>3'</sub>), 3.942 (1H, s, Tyr<sub>α</sub>), 3.886 (1H, d, Phe<sub>α</sub>), 2.58 (1H, m, Tyr<sub>β</sub>), 2.50 (1H, m, Tyr<sub>β</sub>), 2.18 (2H, m, Phe<sub>β</sub>); <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>, δ): 166.13 (s, Tyr<sub>CO</sub>), 166.12 (s, Phe<sub>CO</sub>), 156.07 (s, Tyr<sub>4'</sub>), 136.62 (s, Phe<sub>1'</sub>), 130.75 (s, Tyr<sub>1'</sub>), 129.69 (d, Tyr<sub>2'</sub>), 128.13 (d, Phe<sub>3'</sub>), 127.90 (d, Phe<sub>2'</sub>), 126.38 (d, Phe<sub>4'</sub>), 114.98 (d, Tyr<sub>3'</sub>), 55.68 (d, Tyr<sub>α</sub>), 55.36 (d, Phe<sub>α</sub>), 39.08 (t, Tyr<sub>β</sub>), 38.48 (t, Phe<sub>β</sub>).

**Cyclo(benzyl-Gly) (7):** white powder, C<sub>10</sub>H<sub>10</sub>N<sub>2</sub>O<sub>2</sub>. ESI-MS: *m/z* 213.2 [M+Na]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ, J/Hz): 7.216–7.337 (5H, m, benzyl<sub>Ar</sub>), 4.28 (1H, dd, J = 6, benzyl<sub>α</sub>), 3.38 (1H, m, Gly<sub>α</sub>), 2.89 (1H, m, Gly<sub>α</sub>); <sup>13</sup>C NMR (150 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 174.81 (s, benzyl<sub>CO</sub>), 157.60 (s, Gly<sub>CO</sub>), 135.31 (s, Ar<sub>1'</sub>), 129.15 (d, Ar<sub>2'</sub>), 128.63 (d, Ar<sub>3'</sub>), 127.17 (d, Ar<sub>4'</sub>), 59.70 (d, benzyl<sub>α</sub>), 37.61 (t, Gly<sub>α</sub>).

**Cyclo(Val-Val) (8):** white powder, C<sub>10</sub>H<sub>18</sub>N<sub>2</sub>O<sub>2</sub>. ESI-MS: *m/z* 222 [M+Na +H]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>Cl, δ): 5.923 (1H, s, NH), 4.116 (1H, d, Val<sub>α</sub>), 2.5 (1H, m, Val<sub>β</sub>), 1.25 (3H, d, Val<sub>γ</sub>), 0.9 (3H, d, Val<sub>γ</sub>); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>Cl, δ): 167.42 (s, CO), 59.83 (d, Val<sub>α</sub>), 30.91 (d, Val<sub>β</sub>), 18.26 (q, Val<sub>γ</sub>), 16.27 (q, Val<sub>γ</sub>).

**Cyclo(Gla-Tyr) (9):** white powder, C<sub>11</sub>H<sub>12</sub>N<sub>2</sub>O<sub>3</sub>. ESI-MS: *m/z* 243.7 [M+Na]<sup>+</sup> 219.8 [M-H]<sup>-</sup>. <sup>1</sup>H NMR (600 MHz, DMSO-d<sub>6</sub>, δ, J/Hz): 9.85 (1H, s, OH), 8.059 (1H, d, Tyr<sub>NH</sub>), 7.817 (1H, s, Gly<sub>NH</sub>), 6.915 (2H, d, J = 8.4, Tyr<sub>2'</sub>), 6.636 (2H, d, J = 8.4, Tyr<sub>3'</sub>), 3.954 (1H, d, Tyr<sub>α</sub>), 3.28 (1H, m, Gly<sub>α</sub>), 2.97 (1H, m, Gly<sub>α</sub>), 2.74 (1H, m, Tyr<sub>β</sub>), 2.68 (1H, m, Tyr<sub>β</sub>); <sup>13</sup>C NMR (DMSO-d<sub>6</sub>, 150 MHz): δ 167.38 (s, Tyr<sub>CO</sub>), 165.73 (s, Gly<sub>CO</sub>), 156.27 (s, Tyr<sub>4'</sub>), 131.02 (d, Tyr<sub>2'</sub>), 125.73 (s, Tyr<sub>1'</sub>), 114.97 (d, Tyr<sub>3'</sub>), 55.76 (d, Tyr<sub>α</sub>), 43.66 (t, Gly<sub>α</sub>), 38.17 (t, Tyr<sub>β</sub>).

**Cyclo(Ala-Tyr) (10):** white powder, C<sub>12</sub>H<sub>14</sub>N<sub>2</sub>O<sub>3</sub>. ESI-MS: *m/z* 257.8 [M+Na]<sup>+</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, δ, J/Hz): 6.994 (2H, d, J = 8.4, Tyr<sub>2'</sub>), 6.704 (2H, d, J = 9, Tyr<sub>3'</sub>), 4.22 (1H, m, Tyr<sub>α</sub>), 3.76 (1H, m, Ala<sub>α</sub>), 3.15 (1H, dd, J = 3.6, Tyr<sub>β</sub>), 2.8 (1H, dd, J = 4.8, Tyr<sub>β</sub>), 0.6 (3H, d, Ala<sub>β</sub>); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD, δ): 170.71 (s, Tyr<sub>CO</sub>), 170.63 (s, Ala<sub>CO</sub>), 157.95 (s, Tyr<sub>4'</sub>), 132.75 (d, Tyr<sub>2'</sub>), 127.1 (s, Tyr<sub>1'</sub>), 116.32 (d, Tyr<sub>3'</sub>), 57.6 (d, Tyr<sub>α</sub>), 51.75 (d, Ala<sub>α</sub>), 39.55 (t, Tyr<sub>β</sub>), 20.43 (q, Ala<sub>β</sub>).

**1-(4-Hydroxyphenyl)ethanone (11):** a light green oil, C<sub>8</sub>H<sub>8</sub>O<sub>2</sub>. ESI-MS: *m/z* 159.0 [M+Na]<sup>+</sup>, *m/z* 135.1 [M-H]<sup>-</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ, J/Hz): 7.8 (2H, dd, J = 4.8, Ar-2,2'), 6.8 (2H, dd, J = 4.8, Ar-3,3'), 2.543 (3H, s, CH<sub>3</sub>); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD, CD<sub>3</sub>Cl, δ): 197.87 (s, CO), 161.9 (s, Ar<sub>4'</sub>), 130.82 (d, Ar<sub>2'</sub>), 129.00 (s, Ar<sub>1'</sub>), 115.13 (d, Ar<sub>3'</sub>), 25.95 (t, CH<sub>3</sub>).

**Macrolactin B (12):** a white solid, C<sub>30</sub>H<sub>44</sub>O<sub>10</sub>. ESI-MS: *m/z* 587.76 [M+Na]<sup>+</sup>, *m/z* 600.20 [M+Cl]<sup>-</sup>. <sup>1</sup>H NMR (600 MHz, CD<sub>3</sub>OD, δ, J/Hz): 7.19 (1H, dd, J = 15, H-4), 6.71 (1H, dd, J = 15, H-9), 6.68 (1H, dd, J = 11.4, H-3), 6.23 (1H, dd, J = 15, H-5), 6.16 (1H, dd, J = 11.2, H-17; 1H, dd, J = 15.2, H-10), 6.03 (1H, dd, J = 15, H-18), 5.62 (1H, dd, J = 15, H-8), 5.59 (1H, dd, J = 11.2, H-11), 5.58 (1H, dd, J = 15, H-19), 5.56 (1H, dd, J = 11.4, H-2; 1H, dd, J = 15.2, H-16), 5.0 (1H, m, H-23), 4.49 (1H, dd, H-7), 4.36 (1H, dd, H-15), 4.3 (1H, m, H-1'), 4.1 (1H, m, H-2'), 3.9 (1H, dd, H-13), 3.87 (1H, m, H-6'), 3.34 (1H, m, H-4'), 3.28 (1H, m, H-3'), 3.22 (1H, m, H-5'), 2.4, 2.5 (2H, m, H-12), 2.4, 2.5 (2H, m, H-6), 2.1, 2.2 (2H, m, H-20), 1.6 (2H, m, H-14), 1.6 (2H, m, H-22), 1.5 (2H, m, H-21), 1.2 (3H, d, H-24); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>OD, δ): 167.95 (s, C-1), 145.30 (d, C-3), 142.2 (d, C-5), 135.46 (d, C-16), 134.85 (d, C-19), 133.96 (d, C-8), 131.78 (d, C-18), 131.37 (d, C-17), 130.93 (d, C-4), 129.87 (d, C-10), 129.42 (d, C-9), 129.09 (d, C-11), 117.78 (d, C-2), 101.12 (d, C-1'), 78.26 (d, C-4'), 77.99 (d, C-7), 75.18 (d, C-3'), 72.41 (d, C-5'), 71.88 (d, C-2'), 71.84 (d, C-23), 69.73 (d, C-15), 69.10 (d, C-13), 62.9 (t, C-6'), 43.66 (t, C-14), 41.4 (t, C-6), 36.54 (t, C-12), 35.99 (t, C-22), 32.83 (t, C-20), 25.54 (t, C-21), 20.13 (q, C-24).

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## REFERENCES

1. Hu Yong-mei, Wang Hao, Ye Wen-cai, and Liu Ge, *Zhongguo Yaoke Daxue Xuebao*, **6**, 523 (2005).
2. Cheng Zhi-hong, Wu Tao, and Yu Boyang, *Tianran Chanwu Yanjiu Yu Kaifa.*, **17** (1), 1 (2005).
3. V. Ivanova, U. Graefe, R. Schlegel, B. Schlegel, A. Gusterova, M. Kolarova, and K. Aleksieva, *Biotechnol. Biotechnol. Equip.*, **17** (2), 128 (2003).
4. F. J. Schmitz, D. J. Vanderah, K. H. Hollenbeak, C. E. L. Enwall, V. Gopichand, P. K. Sen Gupta, M. B. Hossain, and D. V. Helm, *J. Org. Chem.*, **48**, 3941 (1983).
5. J. Kang and R. Y. Chen, *Chin. J. Chin. Mater. Med.*, **28** (11), 1038 (2003).
6. H. J. Li, Y. C. Lin, and X. H. Liu, *Acta Sci. Nat. Univ. Suny*, **41** (1), 110 (2002).
7. Sato Nobuhiro and Kato Yukihiro, *J. Heter. Chem.*, **23**, 1677 (1986).
8. Ogura Haruo, Furuhata Kimio, and Furuhata Katsuko, *Chem. Pharm. Bull.*, **23**, 2474 (1975).
9. K. Suzuki, Y. Sasaki, N. Endo, and Y. Mhara, *Chem. Pharm. Bull.*, **29**, 233 (1981).
10. Guangying Chen, Yongcheng Lin, L. L. P. Vrijmoed, and Wang-Fun Fong, *Chem. Nat. Comp.*, **42**, 138 (2006).
11. X. W. Yang, J. S. Wang, Y. H. Wang, H. T. Xiao, X. J. Hu, Sh. Zh. Mu, Y. L. Ma, H. Lin, H. P. He, L. Li, and X. J. Hao, *Planta Med.*, **73**, 496 (2007).
12. K. Gustafson, M. Roman, and William Fenical, *J. Am. Chem. Soc.*, **111**, 75199 (1989).